

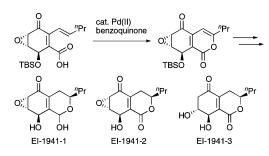
Enantio- and Diastereoselective Total Synthesis of EI-1941-1, -2, and -3, Inhibitors of Interleukin-1 β Converting Enzyme, and **Biological Properties of Their Derivatives**

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The first asymmetric total synthesis of EI-1941–1, -2, and -3, inhibitors of the interleukin-1 β converting enzyme (ICE), has been accomplished, starting from a chiral epoxy iodoquinone 11, a key intermediate in our total synthesis of epoxyquinols A and B. Despite a failure to synthesize the inhibitors by our postulated biosynthetic route, we were able to diastereoselectively synthesize them via an intramolecular carboxypalladation with the key steps being a 6-endo cyclization mode followed by β -hydride elimination. The investigation of the biological properties of EI-1941-1, -2, and -3 and their derivatives disclosed them to be potent and effective ICE inhibitors with less cytotoxicity than EI-1941-1 and -2 in a cultured cell system.

Introduction

Interleukin-1 β (IL-1 β) is an important mediator of pathogenesis of rheumatoid arthritis, septic shock, inflammation, and other physiological conditions.¹ IL-1 β converting enzyme (ICE) is a cysteine protease, which cleaves a biologically inactive 31 kDa precursor to biologically active IL-1 β .² IL-1 β is released from macrophage-like cells in an inflammatory situation, and is the major form of IL-1 in diseases. ICE inhibitors have been shown to prevent inflammation in several acute models,³ suggesting that ICE inhibitors would be useful as antiinflammatory drugs. Recently, Koizumi and co-

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workers have isolated EI-1941-1(1), EI-1941-2(2), and EI-1941-3 (3) from culture broths of *Farrowia* sp., the first two of which selectively inhibit human recombinant ICE activity with IC₅₀ values of 0.086 and 0.006 μ M, respectively, whereas the last is inactive at concentrations up to 10 μ M in an in vitro system.⁴ EI-1941–2 also has weak antimicrobial activities against Gram-positive

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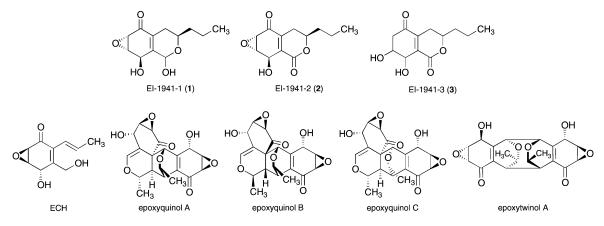


FIGURE 1. Natural products of epoxyquinol monomers and dimers.

bacteria, and moderate activity against *Proteus vulgaris*.⁴ A more potent and effective ICE inhibitor would be desired, and EI-1941–1 and -2 would be suitable lead compounds for the study of the structure–activity relationship.

Structurally, EI-1941-1 and EI-1941-2 have an epoxyquinone core and a side chain. We have been interested in the synthesis and biology of epoxyquinone derivatives such as ECH ((2R,3R,4S)-2,3-epoxy-4-hydroxy-5-hydroxymethyl-6-(1E)-propenylcyclohex-5-en-1one),⁵ an inhibitor of Fas-mediated apoptosis, and its dimer, epoxyquinols A, B, and C, and epoxytwinol A, angiogenesis inhibitors.⁶ At the time that we started this project, the relative and absolute stereochemistries of EI-1941-1, -2, and -3 were not known. As most of the epoxyquinol natural products have a trans relationship between the epoxide and the 4-hydroxy group on the cyclohexenone,⁷ work on a synthetic route by which the two diastereomers (EI-1941-2 and epi-EI-1941-2) can be generated with high optical purity was undertaken in order to determine the relative stereochemistries. As for the absolute stereochemistry, with the structural similarity between EI-1941 and ECH, we tried to synthesize the (1R,5S,6R)-1,6-epoxy-5-hydroxycyclohexenone derivative as our first target.

When we had nearly finished the synthesis of the targeted isomer of EI-1941-2 and its epimer, the absolute and relative stereochemistries of EI-1941-1 and -2 were reported (see Figure 1),⁸ whereas those of EI-1941-3 were not determined because of its low availability from the fermentation broth. Those determina-

tions were made on the basis of the crystallographic analysis of the *p*-bromobenzoyl ester of EI-1941–2 and the chemical correlation between EI-1941–1 and –2. These results indicate that the compounds we have synthesized are opposite enantiomers of the natural EI-1941–2 and its epimer, which was communicated in a previous letter.⁹

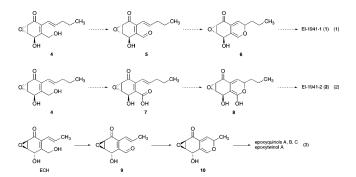
As for the biosynthesis, we postulated the following path: Oxidation of alcohol **4** would afford aldehyde **5**, from which 6π -electrocyclization¹⁰ proceeds to generate 2H-pyran **6**. Hydration and isomerization would afford EI-1941-1, the oxidation of which would provide EI-1941-2 (eq 1). Another possible path involves the oxidation of alcohol **4** to carboxylic acid **7**, from which 6π -electrocyclization proceeds to generate hydroxy-2H-pyran **8** (eq 2). Isomerization would afford EI-1941-2, the reduction of which would provide EI-1941-1. Similar 2H-pyran **10** is a key intermediate of our biomimetic total synthesis of epoxyquinols A, B, and C, and epoxytwinol A, which was generated by the oxidation of ECH followed by the 6π -electrocyclization.^{6c,f,g}

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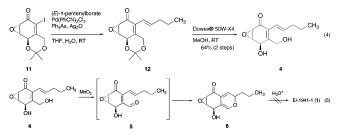
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In this full paper we will describe the highly stereoselective, asymmetric total synthesis of the natural enantiomers of EI-1941–1, -2, and -3 in a full account, including an attempted total synthesis based on a biomimetic route with the theoretical calculation of 6π electrocyclization of dienecarboxylic acid derivatives. We also describe the biological properties of EI-1941–1, -2, and -3 and their derivatives, including the finding of a more superior ICE inhibitor that is less cytotoxic than EI-1941–1 and -2.

Results and Discussion

Synthetic Study Based on Our Postulated Biosynthetic Pathway. As described in the Introduction, EI-1941-1 would be generated by the hydration and isomerization of 2*H*-pyran **6**, and we already found that the similar 2H-pyran 10, generated by the oxidation of ECH, dimerized gradually under neat conditions or in a rather condensed solution (eq 3).^{6c,f,g} We thought the vinyl ether moiety of 6 would react with H₂O before it dimerizes when 2*H*-pyran 6 was treated with acid in a dilute solution. Epoxycyclohexenol 4, the starting material, was synthesized from the chiral iodocyclohexenone 11,^{6c,d,g} an enantiomer of the intermediate of our total synthesis of epoxyquinols, by the Suzuki coupling reaction with (E)-1-pentenylborate¹¹ and Ag₂O in the presence of a catalytic amount of Pd(PhCN)₂Cl₂ and Ph₃As,¹² followed by cleavage of the acetonide on acid treatment (eq 4). 2H-Pyran derivative 6 was isolated after the oxidation of alcohol 4 with MnO₂, followed by 6π -electrocyclization. Though 2Hpyran 6 was treated with several acids such as PPTS, TsOH·H₂O, and CF₃CO₂H in several dilute aqueous solvents, a complex mixture was obtained without isolation of the desired product (eq 5), which prompted us to examine the 6π -electrocyclization of a diene carboxylic acid derivative.



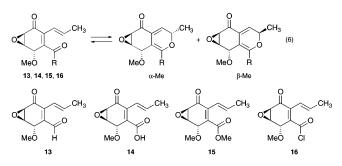
Despite the facile 6π -electrocyclization of dienal 5, 6π electrocyclization has generally been regarded as a

TABLE 1. Calculated TS Energy and Relative Energy between Diene Carbonyl Compound and α - and β -Methyl 2*H*-Pyran Derivatives

entry	starting material	$\begin{array}{c} TS \ energy \\ (kcal \ mol^{-1}) \end{array}$	relative energy ^a (kcal mol ⁻¹)	$isomer^b$
1	13	17.74	-4.16	α
2	13	15.50	-4.18	β
3	14	25.34	+5.78	ά
4	14	8.83	+6.26	β
5	15	25.65	+9.24	ά
6	15	10.23	+8.66	β
7	16	22.50	+1.73	ά
8	16	9.46	+1.58	β

^{*a*} The values are relative energies between the diene carbonyl compound and 2*H*-pyran in the 6π -electrocyclization. ^{*b*} α indicates the α -methyl isomer, whereas β indicates the β -methyl isomer.

difficult reaction.¹⁰ According to the recent theoretical calculation of 2,4-pentadienal, a simple dienal, 6π -electrocyclization is an equilibrium that shifts to the starting material, and 2*H*-pyran is more energetically unstable than the parent aldehyde with a TS energy of 3.44 kcal/ mol versus 21.52 kcal/mol.¹³ In the case of ECH, however, an electron-withdrawing keto group on cyclohexane reduces the TS energy with the stabilization of the 2Hpyran intermediate 10 (eq 3).^{6f} As 6π-oxa-electrocyclization has been investigated only for the diene-aldehydes without a systematic study of that for diene-carboxylic acids or esters, the theoretical calculations for the substrates having a propenyl side chain and functional groups such as aldehyde 13, carboxylic acid 14, methyl ester 15, and acid chloride 16 were performed at the B3LYP/6-31G* level with the program package TITAN 1.0.5 including DFT engine Jaguar 3.5.042.2.14 For all the transition-state searches, vibrational frequencies were computed after completion of the optimization from analytic second derivatives.



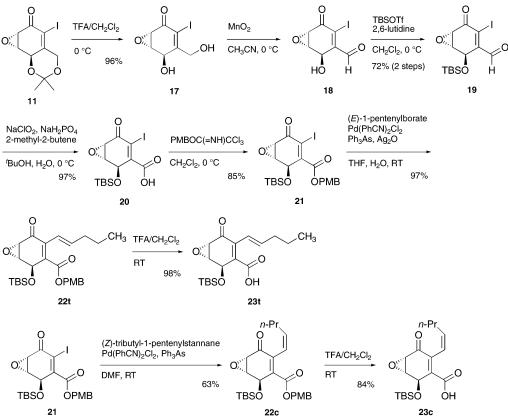
There are two isomers for the 6π -oxa-electrocyclized products such as α - and β -methyl derivatives (eq 6). The transition-state energies for both α - and β -methyl-2*H*pyran derivatives, and the relative energies between diene carbonyl derivatives and 2*H*-pyran derivatives, are calculated with the results summarized in Table 1. We have experimentally demonstrated the facile electrocyclization of diene aldehyde **13**, which is supported by the calculation showing that the 6π -electrocyclized product is more stable than the starting material with the low transition-state energies of 17.74 and 15.50 kcal/mol to α - and β -methyl 2*H*-pyran derivatives, respectively (en-

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SCHEME 1. Synthesis of 23t and 23c



tries 1, 2). Diene carboxylic acid derivatives 14, 15, and 16 showed results different from those of diene aldehyde 13. 6π -Electrocyclization might proceed, judging from the low transition-state energy to the β -methyl isomer, whereas that to the α -methyl isomer is too high for the 6π -electrocyclization to proceed at room temperature. As for the relative energies between the starting materials and the 2*H*-pyran derivatives, 2*H*-pyran derivatives are very unstable except for the acid chloride 16, indicating that the equilibrium shifts mostly to the starting material, and the concentration of the 2*H*-pyran derivatives would be quite low. Even in the case of acid chloride 16, though the 2*H*-pyran derivative is slightly more unstable (1.58 kcal/mol) than the starting material, the equilibrium also shifts to the starting material with the low concentration of 2H-pyran.

Synthesis of Carboxylic Acid Derivatives. With the calculation results in hand, we examined the 6π -oxaelectrocyclization of several derivatives. Before describing the results of 6π -oxa-electrocyclization, we will briefly mention the synthesis of carboxylic acid **23t** (see Scheme 1). Cleavage of the acetonide of the chiral iodocyclohexenone **11** with an acid treatment gave diol **17**. Selective oxidation of the primary alcohol with excess MnO₂ in CH₃CN gave aldehyde **18**; the secondary alcohol was protected by the use of TBSOTf and 2,6-lutidine, affording **19** in 72% yield over two steps. Oxidation of this aldehyde to the carboxylic acid was successfully performed under Kraus' conditions.¹⁵ The carboxylic acid was protected as its *p*-methoxybenzyl ester **21** by the reaction with 4-methoxybenzyl trichloroacetimidate¹⁶ in 85% yield. The introduction of a side chain by the Suzuki coupling reaction with (*E*)-1-pentenylborate and Ag₂O in the presence of a catalytic amount of Pd(PhCN)₂Cl₂ and Ph₃-As afforded **22t** in 97% yield. Acid treatment then gave carboxylic acid **23t** in excellent yield. The isomer with the *Z* side chain, **23c**, was prepared in good yield by the Stille coupling reaction using (*Z*)-tributyl-1-pentenylstannane in the presence of a catalytic amount of Pd-(PhCN)₂Cl₂ and Ph₃As, followed by the acid treatment.

With the starting materials in hand, we investigated the 6π -electrocyclization of dienecarboxylic acid. No reaction proceeded even at reflux in toluene, with the recovery of the starting materials in the cases of carboxylic acid **23t** and ester **22t**. When acid chloride generated from carboxylic acid **23t** with oxalyl chloride and a catalytic amount of DMF was gently heated to 60 °C in CDCl₃, a complex mixture was obtained. Only decomposition occurred when acid chloride was treated with AlCl₃ to generate the acylium ion.

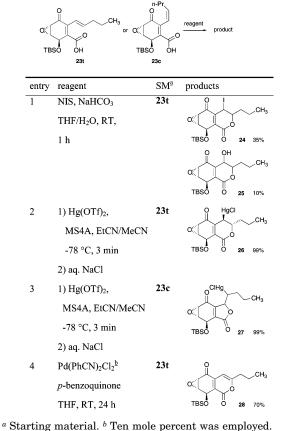
As all our trials using 6π -electrocyclization as a key step were in vain, we pursued another synthetic route using intramolecular carboxymetalation.

Intramolecular Carboxymetalation. Intramolecular addition of the carboxylic acid onto the alkene activated with iodine or metal salts was examined, though diastereoselectivity and alternate reaction modes such as 6-endo or 5-exo are possible problems with this approach (Table 2). In fact, iodolactonization proceeded

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TABLE 2. Intramolecular Cyclization of 23t and 23c



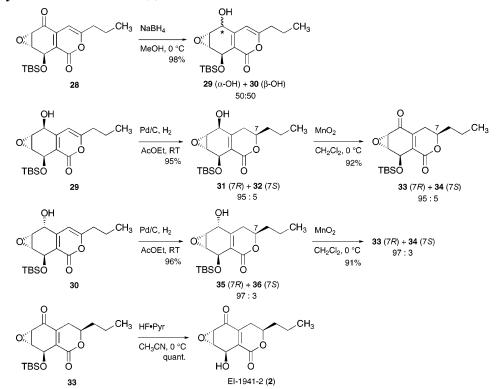
in the 6-endo mode with low yield (entry 1), whereas in the case of carboxymercuration using $Hg(OTf)_{2,}^{17}$ the 6-endo cyclized product was obtained in excellent yield

SCHEME 2. Synthesis of EI-1941-2 (2)

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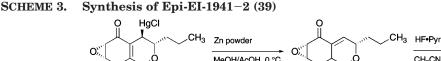
as a single isomer, albeit with the incorrect side-chain relative stereochemistry at C7 in the reaction of *E* isomer **23t** (vide infra, entry 2). Undesired 5-exo cyclization was observed in that of the *Z* isomer **23c** (entry 3). Unlike these unsuccessful results, the 7,8-dihydro-6*H*-iso-chromen-1,5-dione structure **28** was formed when palla-dium(II) was used as a catalyst. That is, when **23t** was treated with *p*-benzoquinone and a catalytic amount of Pd(PhCN)₂Cl₂,¹⁸ carboxypalladation proceeded, followed by the β -hydride elimination, affording **28** in 70% yield (entry 4).

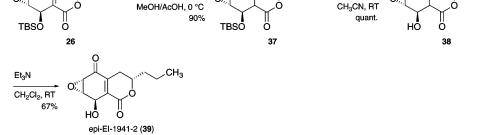
The remaining steps are reduction of the double bond and deprotection (see Scheme 2). Hydrogenation of 28 under an H₂ atmosphere in the presence of Pd/C or Pd- $(OH)_2$ did not proceed. As the keto group might be the cause of this reluctance to undergo hydrogenation, it was reduced with NaBH₄ in MeOH to afford alcohols 29 and 30 in 98% yield and equal amounts, which were separated by column chromatography. The relative stereochemistry is determined by the modified MTPA-ester method¹⁹ of **29**. Hydrogenation of α-alcohol **29** proceeded smoothly and stereoselectively, affording an inseparable mixture of 31 and 32 in excellent yield (95%) and with high diastereoselectivity (95:5). It should be noted that the concentration is important for the diastereoselectivity. Whereas excellent diastereoselectivity (95:5) was obtained at 0.01 M, lower selectivity (87:13) was observed at a higher concentration (0.1 M).^{9,20} The mixture of **31** and 32 was oxidized with MnO₂, affording ketones 33 and 34 in 92% yield (95:5), which were easily separated by thin-layer chromatography (TLC). Though hydrogenation of β -alcohol **30** proceeded slowly, the reduced products 35 and 36 were obtained in 96% yield with the desired isomer stereoselectivity (97:3). In this hydrogenation, the concentration is also crucial. Excellent diastereoselectiv-



IOC Article

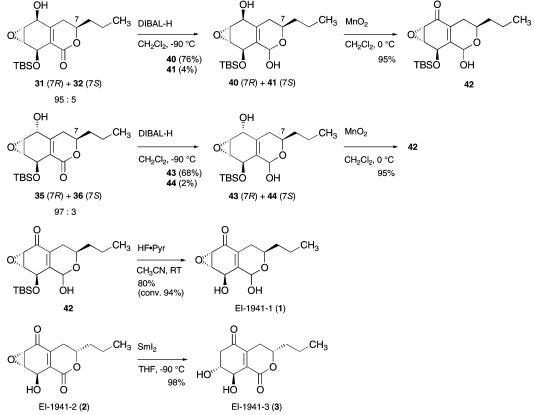
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SCHEME 4.

Synthesis of EI-1941-1 (1) and EI-1941-3 (3)



ity (97:3) is observed at low concentration (0.01 M) in contrast to the lower selectivity (83:17) at higher concentration (0.1 M).9 Oxidation of alcohols 35 and 36 with MnO_2 gave 33 and 34 in 91% yield in a 97:3 ratio, and these were separated by TLC.

Removal of the TBS group of **33** afforded EI-1941-2 (2) quantitatively. Synthetic EI-1941-2 (2) exhibited properties identical to those of the natural product,^{4b,8} including the optical rotation.

epi-EI-1941-2 was also prepared stereoselectively from carboxymercurated derivative 26. Though conventional demercuration using Bu₃SnH in the presence of AIBN²¹ did not work, affording 23t, we found that the treatment of **26** with Zn powder in MeOH and AcOH²² gave β , γ unsaturated lactone 37. After removal of the TBS group, treatment with a catalytic amount of amine isomerized the double bond to provide epi-EI-1941-2 (39) in 67%yield (Scheme 3).

Synthesis of EI-1941-1 and -3. EI-1941-1 was synthesized (see Scheme 4) from α -alcohol **31** and β -alcohol 35, the intermediates of EI-1941-2, as follows: When α -alcohols **31** and **32** (**31**:**32** = 95:5) were treated with DIBAL-H in CH_2Cl_2 at low temperature (-90 °C),

^{(17) (}a) Nishizawa, M.; Takenaka, H.; Nishida, H.; Hayashi, Y. Tetrahedron Lett. 1983, 24, 2581. (b) Nishizawa, M.; Morikuni, E.; Asoh, K.; Kan, Y.; Uenoyama, K.; Imagawa, H. Synlett 1995, 169. (c) Imagawa, H.; Shigaraki, T.; Suzuki, T.; Takao, T.; Yamada, H.; Sugihara, T.; Nishizawa, M. Chem. Pharm. Bull. 1998, 46, 1341. (d) Nishizawa, M.; Kashima, T.; Sakakibara, M.; Wakabayashi, A.; Takahashi, K.; Takao, T.; Imagawa, H.; Sugihara, T. Heterocycles 2000, 54, 629.

^{(18) (}a) Korte, D. E.; Hegedus, L. S.; Wirth, R. K. J. Org. Chem. 1977, 42, 1329. (b) Minami, T.; Nishimoto, A.; Hanaoka, M. Tetrahedron Lett. 1995, 36, 9505.

⁽¹⁹⁾ Ohtani, I.; Kusumi, T.; Kashman, Y.; Kakisawa, H. J. Am. Chem. Soc. 1991, 113, 4092.

⁽²⁰⁾ The reason for the effect of the concentration on the diastereoselectivity is not clear.

⁽²¹⁾ Whitesides, G. M.; San Filippo, J., Jr. J. Am. Chem. Soc. 1970, 92, 6611.

⁽²²⁾ Fieser, L. F.; Fieser, M. Reagents for Organic Synthesis; John Wiley and Sons: New York, 1967; Vol. 1, p 1276.

TABLE 3. Summary of IC_{50} Values of Test Compounds on IL-1 β Secretion from LPS-Stimulated THP-1 Cells and on Cell Viability of THP-1 Cells

compound	${ m IC}_{50} ext{ values}^a ext{ on IL-1}eta \ ext{ secretion } (\mu ext{M})$	${\operatorname{IC}}_{50} \operatorname{values}^b \operatorname{on cell} \ \operatorname{viability} (\mu { m M})$
EI-1941-1(1)	56	>100
EI-1941-2 (2)	15	40
ent-2	10	68
EI-1941-3 (3)	>100	>100
45	>100	>100
ent- 45	74	>100
46	14	20
ent- 46	10	30

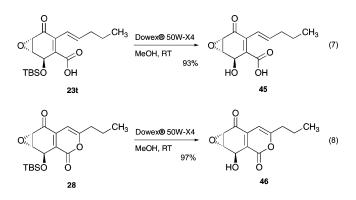
 a Concentration of 50% inhibitory activity on LPS-stimulated IL-1 β secretion. b Concentration of 50% inhibitory activity on cell viability.

lactone was reduced stereoselectively to lactols **40** and **41**, which were separated by TLC (76% **40**, 4% **41**). Oxidation of **40** with MnO₂ gave ketone **42** in excellent yield (95%). β -Alcohols **35** and **36** were also reduced with DIBAL-H to afford lactols **43** and **44**, which were separated by TLC (68% **43**, 2% **44**). Oxidation of **43** gave the same ketone **42** in 95% yield. Deprotection with HF· pyridine afforded EI-1941-1 (**1**) in good yield.

Reduction of the epoxide with Sm_{12}^{23} at low temperature (-90 °C) cleanly converted EI-1941-2 into EI-1941-3 nearly quantitatively. Synthetic EI-1941-1, -2, and -3 exhibited properties identical to those of the natural products, including the optical rotation, which indicate that natural enantiomers were successfully synthesized. Comparison of the optical rotation of EI-1941-3 (synthetic **3**: $[\alpha]^{30}_{\text{D}}$ -88.7, natural **3**:^{4b,8} $[\alpha]^{23}_{\text{D}}$ -87.5) determined its absolute stereochemistry.

Biological Evaluation. We evaluated the effects of EI-1941-1 (1), -2 (2), and -3 (3) and their derivatives on the extracellular release of IL-1 β from THP-1 cells and cell viability in THP-1 cells, with the results summarized in Table 3 and Figure 2. The derivatives examined are *ent*-EI-1941-2 (*ent*-2), hydroxy carboxylic acid **45** and its enantiomer *ent*-**45**, and tetrahydro isocoumarin derivative **46** and its enantiomer *ent*-**46**. Hydroxy carboxylic acid **45** and tetrahydro isocoumarin derivative **46** were easily prepared from **23t** and **28**, respectively, by removal

of the TBS group on acid treatment in MeOH, in good yields (eqs 7 and 8).



Compounds 1 and 2 inhibited IL-1 β secretion in a dosedependent manner; IC_{50} values of 1 and 2 in our assay system were 56 and 15 μ M, respectively. IC₅₀ values of 1 and **2** on cell viability were > 100 and 40 μ M, respectively. Compound 3, which has been reported as an inactive compound in an in vitro system,⁴ was inactive in THP-1 cells. These results indicate that an epoxide ring in **2** is essential for exhibiting the biological activities of 2. Moreover, IC₅₀ values of *ent*-**2** on IL-1 β secretion and cell viability were 10 and 68 μ M, respectively, which was a most effective derivative because the differences in IC_{50} values against IL-1 β secretion and cell viability were greatly significant. The inhibition of IL-1 β secretion by hydroxy carboxylic acids 45 and *ent*-45 was weak (IC₅₀) values of >100 and 74 μ M, respectively), suggesting that a three-fused-ring system was important for the biological activity. Moreover, IC₅₀ values of tetrahydro isocoumarin derivatives **46** and *ent*-**46** on IL-1 β secretion were 14 and 10 μ M, respectively. However, these two compounds 46 and ent-46 also affected the cell viability at IC_{50} values of 20 and 30 μ M, respectively. These results indicate that the enantiomer of EI-1941-2 (ent-2) is a more potent and effective ICE inhibitor than natural EI-1941-2 in a cultured cell system.

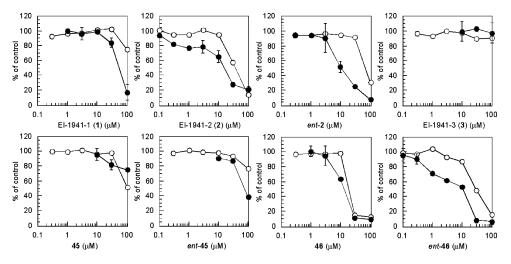


FIGURE 2. Effect of EI-1941-1 (1), -2 (2), and -3 (3) and their derivatives on IL-1 β secretion from LPS-stimulated THP-1 cells and on cell viability of THP-1 cells. Symbols indicate IL-1 β secretion (closed circle) and percentage of viable cells (open circle).

Conclusion

We have accomplished the first asymmetric total synthesis of EI-1941-1, -2, and -3, starting from the chiral epoxy iodoquinone 11, a key intermediate in our total synthesis of epoxyquinols A and B. A key step is the intramolecular, metal-mediated carboxylation of an alkene via 6-endo cyclization, in which Pd(II) gave 2Hpyran-2-one via β -hydride elimination, affording EI-1941–2 after stereoselective hydrogenation. $Hg(OTf)_2$ afforded a carboxymercurated product of a side-chain relative stereochemistry opposite to that of the natural product, leading eventually to epi-EI-1941-2 with high diastereoselectivity. EI-1941-1 was synthesized stereoselectively from the intermediate of EI-1941-2, whereas EI-1941-3 was synthesized in one step from EI-1941-2. By using this asymmetric total synthesis, we determined the absolute stereochemistry of EI-1941-3. The structure-activity relationship of EI-1941-1, -2, and -3 and their synthetic derivatives revealed that an enantiomer of EI-1941-2 is a more potent ICE inhibitor than EI-1941-2, as it is less cytotoxic.

Experimental Section

(1*R*,5*S*,6*R*)-5-Hydroxy-4-hydroxymethyl-3-iodo-7oxabicyclo[4.1.0]hept-3-ene-2-one (17). To a solution of acetonide 11 (100 mg, 0.311 mmol) in CH₂Cl₂ (3.1 mL) was added trifluoroacetic acid (3.1 mL) at 0 °C, and the reaction mixture was stirred for 1 h. The reaction mixture was concentrated under reduced preaaure. The residue was purified by silica gel column chromatography (AcOEt/hexane = 1:1) to afford diol 18 (84 mg, 96%) as a yellow oil: ¹H NMR (400 MHz, CD₃OD) δ 3.63 (1H, dd, J = 1.6, 3.6 Hz), 3.85 (1H, dd, J = 1.4, 3.6 Hz), 4.44 (2H, d, J = 3.2 Hz), 4.98 (1H, br-s); ¹³C NMR (100 MHz, CD₃OD) δ 52.5, 57.4, 64.6, 69.6, 102.1, 163.2, 189.8; FT-IR (neat) ν 3392, 1682, 1591, 1273, 1228, 1080, 1051, 931, 856, 758, 525 cm⁻¹; HRMS (FAB) [M + Na]⁺ calcd for [C₇H₇IO₄ + Na]⁺ 304.9287, found 304.9301; [α]²³_D -31.7 (*c* 0.212, MeOH).

(1S,2S,6R)-2-(tert-Butyldimethylsiloxy)-4-iodo-5-oxo-7oxabicyclo[4.1.0]hept-3-ene-3-carbaldehyde (19). To a solution of diol 17 (81 mg, 0.287 mmol) in CH₃CN (2.9 mL) was added MnO_2 (624 mg, 7.18 mmol) at 0 °C under an argon atmosphere, and the reaction mixture was stirred for 10 min at that temperature. The reaction mixture was filtered through a pad of Celite, and concentrated in vacuo. The residue was filtered through a pad of silica gel (AcOEt/hexane = 1:3) to afford aldehyde 18, and the crude product was used for the next reaction without further purification. To a solution of aldehyde $\mathbf{18}\,(170~\text{mg},\,0.606~\text{mmol})$ and $\text{TBSOTf}\,(481~\text{mg},\,1.82$ mmol) in CH₂Cl₂ (6.1 mL) was added 2,6-lutidine (0.23 mL, 1.94 mmol) at 0 °C, and the mixture was stirred for 1 h under an argon atmosphere. The reaction mixture was quenched with saturated aqueous NH₄Cl, and diluted with AcOEt. The organic phase was washed with saturated aqueous NaCl, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by silica gel column chromatography (AcOEt/hexane = 1:10) to afford siloxy aldehyde 19 (170 mg, 72%, 2 steps) as a yellow oil: ^{1}H NMR (400 MHz, CDCl₃) δ 0.08 (3H, s), 0.22 (3H, s), 0.82 (9H, s), 3.73 (2H, d, J = 1.2 Hz), 5.21 (1H, br-s), 9.79 (1H, s); ¹³C NMR (100 MHz, CDCl₃) δ -4.8, 18.0, 25.6, 51.8, 56.2, 63.5, 117.9, 148.2, 190.6, 197.0; FT-IR (neat) v 2954, 2929, 2858, 1705, $1689, 1581, 1471, 1340, 1255, 1167, 1049, 839, 781, 511 \text{ cm}^{-1};$ HRMS (FAB) $[M + H]^+$ calcd for $C_{13}H_{20}IO_4Si$ 395.0176, found 395.0178; $[\alpha]^{22}_{D}$ -3.2 (c 0.56, MeOH).

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(1S,2S,6R)-2-(tert-Butyldimethylsiloxy)-4-iodo-5-oxo-7oxabicyclo[4.1.0]hept-3-ene-3-carboxylic Acid (20). To a solution of siloxy aldehyde 19 (100 mg, 0.25 mmol), NaH₂PO₄ (40 mg, 0.25 mmol), and 2-methyl-2-butene (0.1 mL, 1.11 mmol) in tert-BuOH (1.8 mL) and H₂O (0.5 mL) was added NaClO₂ (78 mg, 0.86 mmol) at 0 °C, and the reaction mixture was stirred for 1 h under an argon atmosphere. The reaction mixture was concentrated under reduced pressure. The residue was purified by silica gel column chromatography (MeOH/ $CHCl_3 = 1:10$) to afford carboxylic acid **20** (104 mg, 97%) as a colorless oil: ¹H NMR (400 MHz, CDCl₃) δ 0.07 (3H, s), 0.14 (3H, s), 0.84 (9H, s), 3.62 (1H, br-d, J = 3.0 Hz), 3.69 (1H, br-s), 3.79 (1H, br-s), 5.18 (1H, br-s); ¹³C NMR (100 MHz, $\mathrm{CDCl}_3)\,\delta\,-4.6,\,-4.56,\,18.0,\,25.6,\,51.0,\,56.8,\,66.7,\,103.2,\,152.9,$ 170.8, 189.1; FT-IR (neat) v 3199, 2954, 2929, 2858, 1689, 1604, 1389, 1259, 839, 781, 756, 501 cm⁻¹; HRMS (FAB) [M + Na]⁺ calcd for $[C_{13}H_{19}IO_5Si + Na]^+ 432.9944$, found 432.9938; $[\alpha]^{23}D$ +26.1 (c 0.12, MeOH).

(1S,2S,6R)-2-(tert-Butyldimethylsiloxy)-4-iodo-5-oxo-7oxabicyclo[4.1.0]hept-3-ene-3-carboxylic Acid 4-Methoxybenzyl Ester (21). To a solution of carboxylic acid 20 (50 mg, 0.12 mmol) in CH₂Cl₂ was added PMBOC(=NH)CCl₃ (102 mg, 0.24 mmol) at 0 °C, and the reaction mixture was stirred for 1 h under an argon atmosphere. The reaction mixture was quenched with pH 7.0 phosphate buffer, and diluted with AcOEt. The organic phase was washed with saturated aqueous NaCl, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by silica gel column chromatography (AcOEt/hexane = 1:3-1:10) to afford PMB ester 21 (55 mg, 85%) as a yellow oil: ^{1}H NMR (400 MHz, CDCl₃) & 0.01 (3H, s), 0.11 (3H, s), 0.83 (9H, s), 3.62 (1H, dd, J = 1.3, 3.7 Hz), 3.64 (1H, dd, J = 1.0, 3.7 Hz), 3.79 (3H, s), 5.04 (1H, m), 5.21 (2H, dd, J = 11.7, 18.4 Hz), 6.87 (2H, m),7.35 (2H, m); ¹³C NMR (100 MHz, CDCl₃) δ -5.1, -4.6, 17.9, 25.4, 50.9, 55,3, 57.0, 66.8, 68.2, 103.8, 114.0, 126.3, 131.0, 151.3, 160.1, 166.0, 188.6; FT-IR (neat) v 2954, 2858, 1699, 1516, 1241, 1107, 1034, 841, 781 cm⁻¹; HRMS (FAB) [M]⁺ calcd for C₂₁H₂₇IO₆Si 530.0622, found 530.0637; [α]²²_D +19.9 (*c* 0.158, MeOH).

(1S,2S,6R)-2-(tert-Butyldimethylsiloxy)-5-oxo-4-pentyl-7-oxa-bicyclo[4.1.0]hept-3-ene-3-carboxylic Acid 4-Methoxybenzyl Ester (22t). To a solution of PMB ester 21 (48 mg, 0.09 mmol), (E)-1-pentenylborate (16 mg, 0.136 mmol), Ag₂O (33.6 mg, 0.145 mmol), and Ph₃As (2.8 mg, 0.009 mmol) in THF.H₂O (8:1, 0.45 mL) was added Pd(PhCN)₂Cl₂ (1.7 mg, 0.0045 mmol) at room temperature in the dark, and the reaction mixture was stirred for 14 h under an argon atmosphere. The reaction mixture was quenched with saturated aqueous NH₄Cl, and stirred for 30 min at that temperature. The reaction mixture was filtered through a pad of Celite, and the organic materials were extracted with AcOEt. The organic phase was washed with saturated aqueous NaCl, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by silica gel column chromatography (AcOEt/hexane = $1:5 \sim 1:20$) to afford dienone 22t (42 mg, 97%) as a yellow oil: ¹H NMR (400 MHz, CDCl₃) δ 0.00 (3H, s), 0.09 (3H, s), 0.82 (9H, s), 0.84 (3H, t, J = 7.3Hz), 1.32 (2H, sextet, J = 7.3 Hz), 1.90-2.02 (2H, m), 3.54 (1H, d, J = 4.0 Hz), 3.60 (1H, dd, J = 1.9, 4.0 Hz), 3.79 (3H, J)s), 5.10 (2H, d, J = 11.8 Hz), 5.20-5.23 (2H, m), 6.23 (1H, td, J = 6.6, 15.9 Hz), 6.36 (1H, d, J = 15.9 Hz), 6.86–6.88 (2H, m), 7.29–7.32 (2H, m); ¹³C NMR (100 MHz, CDCl₃) δ –4.8, -4.7, 13.7, 17.9, 21.8, 25.5, 35.9, 53.6, 55.3, 56.0, 65.7, 67.2,114.0, 121.7, 127.0, 130.7, 135.5, 136.2, 142.1, 160.0, 167.2, 195.7; FT-IR (neat) v 2956, 2931, 2359, 1716, 1699, 1516, 1244, 1101, 1078, 839, 779 cm⁻¹; HRMS (FAB) [M + H]⁺ calcd for $C_{26}H_{37}O_6Si$ 473.2359, found 473.2385; $[\alpha]^{23}D$ -14.5 (c 0.078, MeOH).

(1S,2S,6R)-2-(*tert*-Butyldimethylsiloxy)-5-oxo-4-pent-1-enyl-7-oxabicyclo[4.1.0]hept-3-ene-3-carboxylic Acid (23t). To a solution of 22t (32 mg, 0.068 mmol) in CH₂Cl₂ (0.7 mL) was added trifluoroacetic acid (0.07 mL) at room temperature, and the reaction mixture was stirred for 1 h. The reaction mixture was concentrated under reduced pressure. The residue was purified by silica gel column chromatography (AcOEt/hexane = 1:1–1:3) to afford carboxylic acid **23t** (23 mg, 98%) as a colorless oil: ¹H NMR (400 MHz, CD₃OD) δ 0.14 (3H, s), 0.21 (3H, s), 0.90 (9H, s), 0.92 (3H, t, J = 7.3 Hz), 1.44 (2H, sextet, J = 7.3 Hz), 1.99–2.21 (2H, m), 3.53 (1H, d, J = 3.9 Hz), 3.72 (1H, dd, J = 3.9 Hz), 5.21 (1H, br-s), 6.32 (1H, td, J = 6.8 Hz, 16.0 Hz), 6.46 (1H, d, J = 16.0 Hz); ¹³C NMR (100 MHz, CD₃OD) δ –4.7, –4.7, 13.7, 17.9, 21.8, 25.5, 36.1, 53.6, 55.7, 65.6, 122.1, 134.5, 137.9, 143.4, 172.6, 196.3; FT-IR (neat) ν 3375, 2929, 2858, 1693, 1680, 1464, 1338, 1099, 781, 741 cm⁻¹; HRMS (FAB) [M + Na]^+ calcd for [C1₁₈H₂₈O₅Si + Na]⁺ 375.1604, found 375.1577; [a]²²_D –61.4 (*c* 0.11, MeOH).

(2S,3S,7R,8S,11R)-3-(tert-Butyldimethylsiloxy)-8-chlormercurio-7-propyl-1,6-dioxatricyclo[8.1.0.0^{4,9}]undec-4en-5,10-dione (26). To a solution of carboxylic acid 23t (21.0 mg, 0.06 mmol) in EtCN (2.1 mL) were added MS4A (6.3 mg, 30 wt %) and Hg(OTf)₂/MeCN (0.25 mL, 0.071 mmol) at -78 °C, and the reaction mixture was stirred for 3 min. The reaction mixture was quenched with saturated aqueous NaH-CO₃ and saturated aqueous NaCl (1:1). The organic materials were extracted with AcOEt, washed with saturated aqueous NaCl, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by silica gel column chromatography (AcOEt/hexane = 1:3) to afford **26** (35.0 mg, 99%) as a colorless solid: ¹H NMR (400 MHz, CDCl₃) δ 0.12 (3H, s), 0.24 (3H, s), 0.87 (9H, s), 0.96 (3H, t, J = 7.3Hz), 1.49-1.70 (2H, m), 1.77-1.93 (2H, m), 2.82 (1H, br-d, J = 11.6 Hz), 3.70 (1H, br-d, J = 3.8 Hz), 3.75 (1H, dd, J = 1.9, 3.8 Hz), 4.55 (1H, ddd, J = 3.5, 7.9, 11.6 Hz), 5.43 (1H, br-s); ¹³C NMR (150 MHz, CDCl₃) δ -4.9, -4.5, 13.7, 18.2, 25.6, 29.7, 38.8, 42.4, 53.1, 55.4, 61.9, 80.6, 136.5, 142.2, 164.2, 196.9; FT-IR (neat) v 2958, 2929, 2858, 2360, 1714, 1680, 1252, 1090, 839, 781 cm⁻¹; HRMS (FAB) [M + H]⁺ calcd for C₁₈H₂₈ClHgO₅-Si 589.1101, found 589.1074; [α]²³_D +48.8 (*c* 0.087, MeOH).

(2R,3S,11S)-3-(tert-Butyldimethylsiloxy)-7-propyl-1,6dioxatricyclo[8.1.0.0^{4,9}]undec-4,7-dien-5,10-dione (28). To a solution of carboxylic acid 23t (100.0 mg, 0.567 mmol) in THF (5.7 mL) was added *p*-benzoquinone (26.2 mg, 2.84 mmol) and Pd(PhCN)₂Cl₂ (12.4 mg, 0.0567 mmol) at room temperature, and the reaction mixture was stirred for 20 h. The reaction mixture was filtered through a pad of Celite, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (AcOEt/hexane = 1:3-1:5) to afford 28 (70.0 mg, 70%) as a yellow oil: ¹H NMR (400 MHz, CDCl₃) & 0.15 (3H, s), 0.26 (3H, s), 0.85 (9H, s), 0.96 (3H, t, J = 7.5 Hz), 1.68 (2H, sextet, J = 7.5 Hz), 2.47 (2H, t, J = 7.5 Hz), 3.63 (1H, br-d, J = 3.9 Hz), 3.77 (1H, dd, J = 2.0, 3.9 Hz), 5.29 (1H, br-s), 6.28 (1H, br-s); ¹³C NMR (100 MHz, $CDCl_3$) δ -4.9, -4.6, 13.5, 20.1, 25.7, 35.8, 52.6, 56.7, 61.9, 98.1, 125.4, 139.6, 139.6, 162.3, 166.9, 192.9; FT-IR (neat) ν 2956, 2929, 2856, 1732, 1705, 1641, 1577, 1464, 1257, 1086, 839, 781 cm⁻¹; HRMS (FAB) [M]⁺ calcd for C₁₈H₂₆O₅Si 350.1550, found 350.1579; $[\alpha]^{23}_{D}$ –14.1 (c 0.17, MeOH).

(2S,3S,10R,11S)-3-(tert-Butyldimethylsiloxy)-10-hydroxy-7-propyl-1,6-dioxatricyclo[8.1.0.04,9]undec-4,7-dien-5-one (29) and (2S,3S,10S,11S)-3-(tert-Butyldimethylsiloxy)-10-hydroxy-7-propyl-1,6-dioxatricyclo[8.1.0.04,9]undec-4,7-dien-5-one (30). To a solution of 28 (43.0 mg, 0.123 mmol) in MeOH (1.3 mL) was added NaBH₄ (14.0 mg, 0.368 mmol) at 0 °C, and the reaction mixture was stirred for 20 min at that temperature. The reaction mixture was quenched with saturated aqueous NH₄Cl. The organic materials were extracted with AcOEt, washed with saturated aqueous NaCl, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by silica gel column chromatography (CHCl₃) to afford 29 and 30 (42.0 mg, 98%, 50:50 diastereoselectivity) as a colorless oil. 29: ¹H NMR (400 MHz, CDCl₃) δ 0.13 (3H, s), 0.25 (3H, s), 0.86 (9H, s), 0.96 (3H, t, J = 7.5 Hz), 3.43 (1H, m), 3.52 (1H, m), 4.55 (1H, br-d, J = 10.2 Hz), 5.23 (1H, d, J = 2.8 Hz), 5.94 (1H, br-s); ¹³C NMR (125 MHz, CDCl₃) δ 13.5, 17.9, 25.7, 29.6, 50.0, 51.3, 62.7, 67.3, 104.7, 117.2, 149.8, 162.5, 166.9; FT-IR (neat) ν 3419, 2927, 2856, 1726, 1651, 1585, 1464, 1254, 1080, 974, 839, 781 cm⁻¹; HRMS (FAB) [M]⁺ calcd for C₁₈H₂₈O₅Si 352.1706, found 352.1680; [α]²³_D +104.1 (c 0.08, MeOH). **30**: ¹H NMR (400 MHz, CDCl₃) δ 0.12 (3H, s), 0.22 (3H, s), 0.85 (9H, s), 0.95 (3H, t, J = 7.4 Hz), 1.67 (2H, sextet, J = 7.4 Hz), 2.44 (2H, t, J = 7.4 Hz), 3.47 (1H, dd, J = 2.5, 4.3 Hz), 3.59 (1H, dd, J = 2.2 Hz), 6.29 (1H, s); ¹³C NMR (100 MHz, CDCl₃) δ -5.0, -4.6, 13.5, 18.1, 20.2, 25.8, 29.7, 35.8, 53.1, 54.4, 62.4, 66.1, 101.4, 117.2, 149.4, 162.3, 165.4; FT-IR (neat) ν 3410, 2929, 2856, 1722, 1645, 1574, 1464, 1252, 1117, 1065, 920, 839, 777 cm⁻¹; HRMS (FAB) [M]⁺ calcd for C₁₈H₂₈O₅Si 352.1706, found 352.1689; [α]²³_D +81.3 (c 0.21, MeOH).

(2S,3S,7S,10R,11S)-3-(tert-Butyldimethylsiloxy)-10-hydroxy-7-propyl-1,6-dioxatricyclo[8.1.0.04,9]undec-4-en-5one (31) and (2S,3S,7R,10R,11S)-3-(tert-Butyldimethylsiloxy)-10-hydroxy-7-propyl-1,6-dioxatricyclo[8.1.0.0^{4,9}]undec-4-en-5-one (32). To a solution of 29 (10.0 mg, 0.0284 mmol) in AcOEt (2.8 mL) was added 10% Pd/C (3.0 mg, 0.0028 mmol) at room temperature, and the reaction mixture was stirred for 3 h under an H₂ atmosphere. The reaction mixture was filtered through a pad of Celite, and concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (AcOEt/hexane = 1:3) to afford an inseparable mixture of 31 and 32 (9.5 mg, 95%, 95:5 diastereoselectivity) as a colorless oil. 31: ¹H NMR (400 MHz, CDCl₃) δ 0.12 (3H, s), 0.22 (3H, s), 0.85 (9H, s), 0.91 (3H, t, J=7.3Hz), 1.37-1.58 (2H, m), 1.74-1.79 (1H, m), 2.29-2.34 (1H, m), 2.38 (1H, dd, J = 4.9, 18.0 Hz), 2.62 (1H, dd, J = 8.4, 18.0 Hz), 3.34-3.36 (1H, m), 3.43-3.45 (1H, m), 4.32 (1H, br-d, J = 8.7 Hz), 4.45-4.51 (1H, m), 5.03 (1H, d, J = 2.4 Hz); ¹³C NMR (100 MHz, CDCl₃) δ –5.0, –4.7, 13.7, 17.9, 18.2, 25.7, 32.0, 36.4, 50.3, 51.8, 62.2, 67.5, 124.3, 148.6, 163.9; FT-IR (neat) v 3423, 2958, 2931, 2858, 1716, 1254, 1084, 868, 839, 779 cm⁻¹; HRMS (FAB) $[M + H]^+$ calcd for $C_{18}H_{31}O_5Si$ 355.1941, found 355.1948.

(2S.3S.7S.11R)-3-(tert-Butyldimethylsiloxy)-7-propyl-1,6-dioxatricyclo[8.1.0.0^{4,9}]undec-4-en-5,10-dione (33). To a solution of 31 and 32 (10.0 mg, 0.0283 mmol) in CH_2Cl_2 (1.0 mL) was added MnO₂ (61.3 mg, 0.705 mmol) at 0 °C under an argon atmosphere, and the reaction mixture was stirred for 1 h at that temperature. The reaction mixture was filtered through a pad of Celite, and concentrated in vacuo. The residue was purified by preparative thin-layer chromatography (AcOEt/ hexane = 1:5) to afford 33 (9.2 mg, 92%; 95:5 diastereoselectivity) as a colorless oil: ¹H NMR (400 MHz, $CDCl_3$) δ 0.14 (3H, s), 0.45 (3H, s), 0.85 (9H, s), 1.33-1.58 (3H, m), 1.81-1.72 (1H, m), 2.52 (1H, dd, J = 4.6, 18.1 Hz), 2.63 (1H, dd, J)= 7.9, 18.1 Hz), 3.56 (1H, br-d, J = 3.9 Hz), 3.71 (1H, dd, J = 1.9, 3.9 Hz), 4.48-4.54 (1H,m), 5.16 (1H, br-s); ¹³C NMR (100 MHz, CDCl₃) δ -4.8, -4.75, 13.7, 18.1, 18.2, 25.7, 29.7, 36.2, 52.3, 56.4, 61.9, 135.8, 139.5, 163.5, 194.4; FT-IR (neat) v 2927, $2854, 1726, 1695, 1464, 1252, 1240, 1101, 1084, 839, 781 \text{ cm}^{-1};$ HRMS (FAB) $[M+H]^+$ calcd for $C_{18}H_{29}O_5Si\ 353.1784,$ found 353.1782; $[\alpha]^{22}_{D}$ –10.5 (c 0.12, MeOH).

(2*R*,3*S*,7*S*,11*R*)-3-Hydroxy-7-propyl-1,6-dioxatricyclo-[8.1.0.0^{4,9}]undec-4-en-5,10-dione (EI-1941-2 (2)). To a solution of 33 (16.6 mg, 0.0471 mmol) in CH₃CN (1.9 mL) was added HF·Pyr (0.5 mL) at 0 °C, and the reaction mixture was stirred for 4 h at that temperature. The reaction mixture was quenched with saturated aqueous NaHCO₃. The organic materials were extracted with AcOEt, washed with saturated aqueous NaCl, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (AcOEt/hexane = 1:1) to afford EI-1941-2 (2) (11.2 mg, quant.) as a colorless powder: ¹H NMR (400 MHz, CD₃CN) δ 0.93 (3H, t, J = 7.3 Hz), 1.35–1.51 (2H, m), 1.57–1.66 (1H, m), 1.69–1.78 (1H, m), 2.46 (1H, ddd, J = 1.0 9.7, 18.1 Hz), 2.54 (1H, ddd, J = 1.3, 4.6, 18.1 Hz), 3.56 (1H, dd, J = 1.0, 3.7 Hz), 3.85 (1H, dd,
$$\begin{split} J &= 1.6, \, 3.7 \,\, \text{Hz}), \, 4.46 - 4.53 \,\, (1\text{H, m}), \, 4.97 \,\, (1\text{H, br-s}); \, ^{13}\text{C NMR} \\ (100 \,\, \text{MHz}, \, \text{CD}_3\text{CN}) \,\, \delta \,\, 12.5, \,\, 17.3, \,\, 25.3, \,\, 35.6, \,\, 51.8, \,\, 55.8, \,\, 60.4, \\ 77.1, \,\, 135.1, \,\, 140.0, \,\, 163.7, \,\, 193.9; \,\, \text{FT-IR} \,\, (\text{neat}) \,\, \nu \,\, 3448, \,\, 2960, \\ 2873, \,\, 1716, \,\, 1695, \,\, 1417, \,\, 1244, \,\, 1124, \,\, 1099, \,\, 1041 \,\, \text{cm}^{-1}; \,\, \text{HRMS} \\ (\text{FAB}) \,\, [\text{M} + \text{H}]^+ \,\, \text{calcd for} \,\, \text{C}_{12}\text{H}_{15}\text{O}_5 \,\, 239.0920, \,\, \text{found} \,\, 239.0932; \\ [\alpha]^{22}_{\text{D}} \,- 299.9 \,\, (c \,\, 0.48, \,\, \text{MeOH}). \end{split}$$

(2S,3S,7R,11R)-3-(tert-Butyldimethylsiloxy)-7-propyl-1,6-dioxatricyclo[8.1.0.04,9]undec-8-en-5,10-dione (37). To a solution of 26 (5.0 mg, 0.009 mmol) in MeOH (0.1 mL) were added zinc powder (2.8 mg, 0.045 mmol) and AcOH (0.002 mL) at 0 °C, and the reaction mixture was stirred for 10 min. The reaction mixture was quenched with saturated aqueous NaH-CO₃. The reaction mixture was filtered through a pad of Celite, and the organic materials were extracted with AcOEt. The organic phase was washed with saturated aqueous NaCl, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (AcOEt/hexane = 1:5) to afford **37** (2.7 mg, 90%) as a colorless oil: ¹H NMR (400 MHz, CDCl₃) δ 0.07 (1H, s), 0.12 (1H, s), 0.78 (1H, s), 0.93 (3H, t, J = 7.2 Hz), 1.39-1.53 (2H, m), 1.66 - 1.76 (2H, m), 3.47 (1H, d, J = 4.1 Hz), 3.52(1H, q, J = 3.0 Hz), 3.61 (1H, t, J = 4.1 Hz), 5.06 (1H, t, J =3.0 Hz), 7.05 (1H, t, J = 3.0 Hz); ¹³C NMR (100 MHz, CDCl₃) $\delta - 4.1, -4.8, 14.0, 18.2, 25.9, 38.8, 40.7, 54.8, 56.6, 67.5, 79.5,$ 127.9, 135.1, 168.3, 190.7; FT-IR (neat) v 2927, 2856, 1743, 1709, 1655, 1464, 1389, 1117, 839, 781 cm⁻¹; HRMS (FAB) $[M + H]^+$ calcd for $C_{18}H_{29}O_5Si 353.1784$, found 353.1791; $[\alpha]^{23}D_{-1}$ -31.6 (c 0.113, MeOH).

(2R,3S,7R,11R)-3-Hydroxy-7-propyl-1,6-dioxatricyclo-[8.1.0.0^{4,9}]undec-8-en-5,10-dione (38). To a solution of 37 (3.5 mg, 0.01 mmol) in CH₃CN (0.2 mL) was added HF·Pyr (0.05 mL) at room temperature, and the reaction mixture was stirred for 6 h at that temperature. The reaction mixture was quenched with saturated aqueous NaHCO₃. The organic materials were extracted with AcOEt, washed with saturated aqueous NaCl, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (AcOEt/hexane = 1:1) to afford **38** (2.4 mg, quant.) as a colorless oil: ¹H NMR (400 MHz, CDCl₃) δ 0.95 (3H, t, J = 7.4 Hz), 1.38–1.60 (2H, m), 1.67–1.79 (2H, m), 3.52–3.55 (2H, m), 3.79 (1H, dd, J = 3.8, 3.6 Hz), 5.14 (1 H, dd, J = 3.1, 3.6 Hz), 7.13 (1 H, t, J = 3.5 Hz)Hz); ¹³C NMR (100 MHz, CDCl₃) δ 13.6, 18.1, 37.0, 41.1, 54.0, 55.3, 65.0, 79.6, 127.3, 135.9, 169.0, 189.7; FT-IR (neat) v 2956, 2927, 2854, 1743, 1709, 1655, 1250, 1117, 1061, 839, 781 $\rm cm^{-1}$ HRMS (FAB) $[M + H]^+$ calcd for $C_{12}H_{15}O_5$ 239.0920, found 239.0909; $[\alpha]^{32}_{D}$ +37.4(c 0.113, MeOH).

(2R,3S,7R,11R)-3,5-Dihydroxy-7-propyl-1,6-dioxatri $cyclo[8.1.0.0^{4,9}] undec-4\text{-}en-10\text{-}one \ (epi-EI-1941-2 \ (39)). \ To$ a solution of **38** (5.8 mg, 0.024 mmol) in CH₂Cl₂ (0.24 mL) was added Et₃N (1.7 μ L, 0.012 mmol) at room temperature, and the reaction mixture was stirred for 2 h. The reaction mixture was quenched with buffer, and diluted with AcOEt. The organic phase was washed with saturated aqueous NaCl, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (AcOEt/hexane = 1:1) to afford epi-EI-1941-2 (**39**) (3.5 mg, 67%) as a colorless powder: ¹H NMR (400 MHz, CD₃CN) δ 0.95 (3H, t, J = 7.3 Hz), 1.55–1.38 (2H, m), 1.77– 1.62 (2H, m), 2.88 (1H, dd, J = 3.4, 18.1 Hz), 3.59 (1H, dd, J = 1.0, 3.6 Hz), 3.84 (1H, dd, J = 1.5, 3.6 Hz), 4.35-4.42 (1H, m), 5.11 (1H, br-s); ¹³C NMR (100 MHz, CD₃CN) δ 14.1, 18.7, 26.3, 36.3, 53.3, 57.0, 61.3, 78.7, 136.5, 141.3, 165.5, 194.4; FT-IR (neat) v 3438, 2958, 2871, 1716, 1697, 1417, 1124, 1113, 1246, 1041 cm⁻¹; HRMS (FAB) $[M + H]^+$ calcd for $C_{12}H_{15}O_5$ 239.0920, found 239.0922; $[\alpha]^{23}$ _D -29.5 (*c* 0.087, MeOH).

(2R,3R,7R,10S,11R)-3-(tert-Butyldimethylsiloxy)-7-propyl-1,6-dioxatricyclo[8.1.0.0^{4,9}]undec-4-en-5,10-diol (40). To a solution of 31 and 32 (9.0 mg, 0.0254 mmol) in CH₂Cl₂ (0.9 mL) was added a hexane solution of DIBAL-H (0.94 M, 0.09 mL, 0.0787 mmol) at -90 °C under an argon atmosphere, and the reaction mixture was stirred for 20 min at that

temperature. The reaction mixture was quenched with saturated aqueous Rochelle salt. The organic materials were extracted with AcOEt, washed with saturated aqueous NaCl, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (AcOEt/hexane = 1:3) to afford 40 (6.9 mg, 80%, 95:5 diastereoselectivity) as a colorless oil: ¹H NMR (400 MHz, CDCl₃) δ 0.14 (3H, s), 0.17 (3H, s), 0.89 (9H, s), 0.91 (3H, t, J = 7.2 Hz), 1.32–1.60 (2H, m), 1.80 (1H, dd, J = 1.9, 15.5 Hz), 1.88 (1H, br-d, J = 11.1 Hz), 2.32 (1H, dd, J = 11.1, 17.4 Hz, 2.62 (1H, br-d, J = 5.3 Hz), 3.23–3.24 (1H, m), 3.38–3.39 (1H, m), 3.93 (1H, ddt, J = 4.3, 7.5, 11.1 Hz), 4.18 (1H, br-d, J = 9.9 Hz), 4.59 (1H, br-s), 5.37 (1H, d, J = 4.9 Hz); ¹³C NMR (100 MHz, CDCl₃) δ -4.9, -4.5, 14.0, 18.0, 18.5, 25.8, 33.3, 37.2, 52.5, 52.7, 63.3, 65.8, 67.4, 88.8, 129.1, 131.6; FT-IR (neat) v 3410, 2956, 2929, 2858, 2364, 2341, 1259, 1082, 1059, 1003, 974, 837, 779 cm⁻¹; HRMS (FAB) [M]⁺ calcd for $C_{18}H_{32}O_5Si$ 356.2019, found 356.2004; $[\alpha]^{22}D - 7.13$ (c 0.70, MeOH).

(2R,3R,7R,11S)-3-(tert-Butyldimethylsiloxy)-5-hydroxy-7-propyl-1,6-dioxatricyclo[8.1.0.0^{4,9}]undec-4-en-10-one (42). To a solution of **40** (10.5 mg, 0.0295 mmol) in CH₂Cl₂ (1.0 mL) was added MnO₂ (64.0 mg, 0.736 mmol) at 0 °C under an argon atmosphere, and the reaction mixture was stirred for 1 h at that temperature. The reaction mixture was filtered through a pad of Celite, and concentrated in vacuo. The residue was purified by preparative thin-layer chromatography (AcOEt/ hexane = 1:5) to afford 42 (10.0 mg, 95%; 95:5 diastereoselectivity) as a colorless oil: ¹H NMR (400 MHz, CDCl₃) δ 0.17 (3H, s), 0.19 (3H, s), 0.89 (9H, s), 0.91 (3H, t, J = 7.3 Hz), 1.38–1.61 (2H, m), 2.06 (1H, dd, J = 10.8, 17.6 Hz), 2.18 (1H, br-d, J = 17.6 Hz), 2.80 (1H, br-s), 3.47 (1H, dd, J = 1.0, 3.6Hz), 3.61 (1H, dd, J = 1.0, 3.6 Hz), 3.86–3.92 (1H, m), 4.82 (1H, br-s), 5.56 (1H, s); ¹³C NMR (100 MHz, CDCl₃) δ -4.8, -4.3, 13.9, 18.0, 18.4, 25.6, 27.7, 37.1, 52.6, 57.6, 63.3, 66.3,88.0, 129.8, 146.0, 193.4; FT-IR (neat) v 3431, 2956, 2931, 2860, 1684, 1464, 1259, 1074, 866, 739 cm⁻¹; HRMS (FAB) [M + H]⁺ calcd for $C_{18}H_{32}O_5Si$ 356.2019, found 356.1990; $[\alpha]^{21}D$ -137.2 (c 0.593, MeOH).

(2R,3R,7R,11S)-3,5-Dihydroxy-7-propyl-1,6-dioxatricyclo[8.1.0.0^{4,9}]undec-4-en-10-one (EI-1941-1 (1)). To a solution of 42 (6.0 mg, 0.0169 mmol) in CH₃CN (0.5 mL) was added HF·Pyr (0.05 mL) at room temperature, and the reaction mixture was stirred for 4 h. The reaction mixture was quenched with saturated aqueous NaHCO₃. The organic materials were extracted with AcOEt, washed with saturated aqueous NaCl, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (AcOEt/hexane = 1:1) to afford EI-1941-1 (1) (3.2 mg, 94% based on conversion) as a brownish oil: $\,^1\!\mathrm{H}$ NMR (400 MHz, CD_3CN) δ 0.92 (1H, t, J = 7.2 Hz), 1.35 - 1.54 (4H, m), 1.96 (1H, br-dd, J = 11.1, 17.7Hz), 2.09 (1H, ddd, J = 1.8, 3.2, 17.7 Hz), 3.43 (1H, dd, J = 1.0, 3.7 Hz), 3.74 (1H, dd, J = 1.3, 3.7 Hz), 3.85 (1H, m), 4.59(1H, br-s), 5.51 (1H, s); $^{13}\mathrm{C}$ NMR (100 MHz, CD₃CN) δ 14.2, 19.2, 28.3, 37.9, 53.4, 57.7, 62.9, 66.3, 88.2, 129.9, 148.3, 194.9; FT-IR (neat) v 3419, 2960, 2933, 2873, 1682, 1456, 1281, 1028, 874, 725 cm⁻¹; HRMS (FAB) $[M + H]^+$ calcd for $C_{12}H_{17}O_5$ 241.1076, found 241.1075; $[\alpha]^{20}$ _D -188.8 (c 0.16, MeOH).

(3*R*,7*R*,8*R*)-7,8-Dihydroxy-3-propylisochromene-1,5-dione (EI-1941–3 (3)). To a solution of EI-1941–2 (2) (4.6 mg, 0.0193 mmol) in THF (0.39 mL) was added a THF solution of SmI₂ (0.1 M, 0.58 mL, 3.0 mmol) at -90 ° C under an argon atmosphere, and the reaction mixture was stirred for 20 min at that temperature. The reaction mixture was quenched with pH 7.0 phosphate buffer, and diluted with AcOEt. The organic phase was washed with saturated aqueous NaCl, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (AcOEt/hexane = 1:0) to afford EI-1941–3 (3) (4.5 mg, 98%) as a reddish oil: ¹H NMR (400 MHz, CD₃CN) δ 0.94 (3H, t, J = 7.3 Hz), 1.35–1.56 (1H, m), 1.60–1.69 (1H,

m), 1.71–1.80 (1H, m), 2.26 (1H, br-dd, J = 11.3, 18.2 Hz), 2.51 (1H, dd, J = 4.0, 16.7 Hz), 2.76 (1H, ddd, J = 1.4, 3.9, 18.2 Hz), 2.93 (1H, dd, J = 3.0, 16.7 Hz), 3.38 (1H, br-s), 3.76 (1H, br-s), 4.22 (1H, q, J = 3.2 Hz), 4.42–4.49 (1H, m); ¹³C NMR (100 MHz, CD₃CN) δ 14.1, 18.9, 26.3, 37.4, 41.7, 66.7, 70.5, 79.0, 136.9, 143.6, 167.0, 197.7; FT-IR (neat) ν 3419, 2960, 2933, 2873, 1716, 1693, 1410, 1230, 1024 cm⁻¹; HRMS (FAB) [M + H]⁺ calcd for C₁₂H₁₇O₅ 241.1076, found 241.1076; [α]³⁰_D –88.7 (c 0.28, MeOH).

(1S,2S,6R)-2-Hydroxy-5-oxo-4-pent-1-enyl-7-oxabicyclo-[4.1.0]hept-3-ene-3-carboxylic Acid (45). To a solution of 23t (5.0 mg, 0.014 mmol) in MeOH (0.05 mL) was added Dowex 50W-X4 (10.0 mg), and the mixture was stirred at room temperature for 24 h. The reaction mixture was filtered, and the filtrate was concentrated in vacuo. The residue was purified by preparative thin-layer chromatography (MeOH/ $CHCl_3 = 1:10$) to afford carboxylic acid 45 (2.3 mg, 93%) as a colorless oil: ¹H NMR (400 MHz, CD₃OD) δ 0.92 (3H, t, J=7.3 Hz), 1.45 (2H, sextet, J=7.3 Hz), 2.11 (2H, q, J=6.9Hz), 3.54 (1H, d, J = 3.9 Hz), 3.77 (1H, d, J = 3.9 Hz), 5.00(1H, br-s), 6.33 (1H, dt, J = 6.9, 16.1 Hz), 6.45 (1H, d, J =16.1 Hz); ¹³C NMR (100 MHz, CD₃OD) & 13.9, 23.1, 36.9, 55.0, 57.3, 65.9, 123.5, 134.1, 141.3, 143.0, 174.6, 197.2; FT-IR (neat) ν 3342, 2960, 2925, 2873, 2854, 2360, 1695, 1633, 1576, 1261, 1041, 970, 739 cm⁻¹; HRMS (FAB) $[M + H]^+$ calcd for $[C_{12}H_{14}O_5 + H]^+$ 239.0919, found 239.0934; $[\alpha]^{25}D - 73.8$ (c 0.32, MeOH)

(2*R*,3*S*,11*S*)-3-Hydroxy-7-propyl-1,6-dioxatricyclo-[8.1.0.0^{4,9}]undec-4,7-dien-5,10-dione (46). To a solution of 28 (3.6 mg, 0.0074 mmol) in MeOH (0.05 mL) was added Dowex 50W-X4 (7.2 mg), and the mixture was stirred at room temperature for 24 h. The reaction mixture was filtered, and the filtrate was concentrated in vacuo. The residue was purified by preparative thin-layer chromatography (AcOEt/ hexane = 1:5) to afford 46 (1.7 mg, 97%) as a yellow oil: ¹H NMR (400 MHz, CDCl₃) δ 0.96 (3H, t, J = 7.5 Hz), 1.67 (2H, sextet, J = 7.5 Hz), 2.50 (2H, t, J = 7.5 Hz), 3.66 (1H, d, J =3.5 Hz), 3.94 (1H, dd, J = 1.4, 3.5 Hz), 5.28 (1H, br-s), 6.38 (1H, s); ¹³C NMR (100 MHz, CDCl₃) δ 13.4, 20.2, 35.8, 52.8, 56.6, 61.4, 98.2, 125.7, 139.1, 163.4, 166.9, 191.4; FT-IR (neat) ν 3431, 2964, 2931, 2875, 1722, 1705, 1641, 1577, 1045, 852, 760 cm⁻¹; HRMS (FAB) [M]⁺ calcd for $C_{12}H_{12}O_5$ 236.0685, found 236.0664; [α]²⁴_D =96.1 (*c* 0.43, MeOH).

Measurement of Interleukin-1 β) Secretion. THP-1 cells were suspended in RPMI1640 medium (Sigma, St. Louis, MO) supplemented with 10% fetal bovine serum, and seeded on 48well plates (5 × 10⁴ cells/well). The cells were differentiated with 30 nM of pharbol-12-myristate-13-axetate (PMA) for 72 h. After the plate was rinsed with serum-free RPMI1640 medium to remove unadherent cells, adherent cells were stimulated with 100 µg/ml of lipopolysaccharide (LPS; Sigma) for 4 h in the presence of various concentrations of test compounds. The culture media were harvested, and mature IL-1 β was measured by an ELISA method using an IL-1 β assay kit (Amersham Biosciences, Tokyo, Japan).

Measurement of Cell Viability. THP-1 cells $(2.5 \times 10^4 \text{ cells/well})$ were differentiated with 30 nM PMA as described above. The differentiated cells were then treated with test compounds for 4 h. The cell number was evaluated by the subsequent color reaction. WST-8 solution 2-(2-methoxy-4-nitrophenyl)-3-(4-nitrophenyl)-5-(2,4-disulfophenyl)-2H-tetrazolium, monosodium salt (Nacalai tesque, Kyoto, Japan), was added to the medium, and the cells were further incubated for 3 h at 37 °C. The absorbance (A_{450}) of each well was measured using a plate reader (Wallac 1420 multilabel counter; Amersham Biosciences). Cell viability (%) was calculated as (experimental absorbance – background absorbance)/ (control absorbance – background absorbance) $\times 100$.

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Supporting Information Available: Copies of ¹H and ¹³C NMR and IR spectra of all new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

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